

MOLLUSCICIDAL EFFICACY AND TOXICITY OF SOME PESTICIDES UNDER LABORATORY AND FIELD CONDITIONS

[56]

Abdel-Halim¹, K.Y.; R.K. Abou-El Khear² and A.A. Hussein¹

ABSTRACT

Laboratory and field trials were carried out at Koom Hamada district and Etay El-baroud research station during 2004/ 2005 season to clarify the molluscicidal activities of the following: indoxacarb, lufenuron, *Bacillus thuringiensis*, *kurestaci* (*Bt.*) and methomyl against glassy clover snails *Monacha cartusiana* (Müller). Results indicated that all tested pesticides decreased snails population compared with control. Under the field conditions, the efficiency of the tested compounds were 98.0, 93.4, 93 and 71.58% for methomyl, lufenuron, indoxacarb and *Bt.* after 5, 6, 13, and 28 days of treatment, respectively. Moreover the effect of LC₅₀ and 0.5 LC₅₀ of these compounds were investigated on some biochemical parameters *in vivo*. The activities of acetylcholinesterase (AChE), alanine aminotransferases (ALT), aspartate aminotransferases (AST) and protein contents at different time intervals were evaluated. The specific activity of AChE reached 0.009, 0.002, 0.006 and 0.001 µmole/ mg protein/ min for indoxacarb, lufenuron, *Bt.* and methomyl, respectively, after one day of treatment with (LC₅₀ for each compound). In general, *Bt.* and indoxacarb caused slight inhibition on AChE enzyme. All treatments decreased ALT and AST enzyme activity in the tested animals. However, *Bt.* showed slight effect on the activity of aminotransferases enzymes.

Keywords: Snails, Molluscs, Pesticides, Toxicity, Enzymes.

INTRODUCTION

Mollusks are a large and diverse group of animals distributed worldwide (El-Okda, 1980; Godan, 1983 and Nakhla *et al* 1993). Terrestrial gastropods (mollusks: snails and slugs) are being abundantly distributed in the North coast, new rec-

laimed lands and addition to delta region of Egypt (Kasaab and Daoud, 1964); (El-Okda and Khalil, 1981) and (Abou-Baker, 1997). Also, snails are becoming serious agricultural animal pests in Egypt, especially in the Northern coastal areas. Their damage to ornamental plants resembles that done by caterpillars or

1- Central Agric Pesticide Laboratory (CAPL) ARC, Etay El-Baroud, Egypt

2- Plant Protection Res. Ins., Agric Res. Center (ARC), International Potato Center (CIP), Ministry of Agriculture, Kafr El-Zayat, Egypt

(Received April 18, 2006)

(Accepted July 24, 2006)

wireworms. The brown garden snail (*Helix aspersa*), is the common snail causing problems in California gardens; it was introduced from France during the 1850s for use as food. Snails and slugs move by gliding along on a muscular "foot". Snails and slugs feed on a variety of living plants as well as on decaying plant matter. On plants, they chew irregular holes with smooth edges in leaves and can clip succulent plant parts (Flint, 1998). They can also chew fruit and young plant bark. A good snail and slug management program relies on a combination of methods. The first step is eliminate, all places where snails or slugs can hide during the day. Boards, stones, debris, weedy areas around tree trunks, leafy branches growing close to the ground, and dense ground covers such as ivy are ideal sheltering spots safe (Salgado, 1990). Synthetic molluscicides are still considered the most effective measures for the control of snails. Lufenuron is a new insecticide being effective against resistant pests to organophosphates and pyrethroids. It was soft on adult beneficial and predatory mites, safe on a wide range of crops and suitable for integrated pest management (Harder et al 1996). One of the most promising biological control approach that has received attention of many scientists is the development of *Bacillus thuringiensis* (*Bt.*) toxins as insecticides (Belfore et al 1994). Dipel - 2X is one of the biological insecticides containing proteins that are highly toxic to insects. Indoxacarb is the common name proposed for the S-isomer of oxadiazine derivative, which is insecticidal active isomer. This compound represents a new class of compounds has broad-spectrum insecticidal activity and yet be environmentally safe (Salgado, 1990).

The present study was aimed to evaluate the efficacy of some safe pesticides such as indoxacarb, *Bacillus thuringiensis* (*Bt.*) and lufenuron as molluscicides compared with methomyl under field conditions, and examined their effects on some biochemical parameters in the laboratory.

MATERIAL AND METHODS

I- Field Studies

The field study was conducted in clover fields at Koom Hamada district, El-Behera governorate during May, 2005. The treatments were done with 0.5 LC₅₀ and LC₅₀ which cited from Abou- El Khear et al (2005). The LC₅₀ values were 6.7, 6.4, 7.0, and 5.9 x10⁴ ppm for indoxacarb (Avanet® 15% SC) methyl (s)-N- [7-chloro-2, 3, 4, a, 5-tetrahydro-4a (methoxy carbonyl) indeno [1,2-e] [1, 3, 4] oxadiazin-2-yl carbonyl]-4- (trifluoromethoxy) carbonilate; lufenuron (Match® 5% EC): N-[[[2, 5-dichloro-4-(1, 1,2,3,3,3, -hexafluoro-propoxy)-(phenyl) amino] carbonyl]-2, 6-difluorobe-zamide (CA); Dipel® -2X 10.3% WP: *Bacillus thuringiensis*, *kurstaki*, and methomyl (Lannate® 90% SP): S-methyl-N- (methylcarbamoyl) oxyl- thioacetimidate, respectively. The experiments were carried out in plots of ¼ feddan each (1000m²). Each treatment was replicated five times in random blocks. Bait were distributed before sunset using twelve bait stations in different areas of the plot. Wheat bran was kept in bait stations for three days, and then the poisoned baits were added in 200 gm each. The bait stations were inspected and replenished on each subsequent two days until the cessation of the snails' activity, nearly for one

month. Daily bait consumption and dead animals were counted and removed from each plot. Daily observation and snails traps in this area showed that, *Monacha cartusiana* was the most common and important species. Mortality percentage was calculated and the molluscicidal potency expressed as population reduction was measured for all tested compounds according to Henderson and Tilton equation (1955):

$$\text{Reduction \%} = 1 - \left\{ \frac{\text{No. of control before baiting}}{\text{No. of control after baiting}} \times \frac{\text{No. of treatment after baiting}}{\text{No. of treatment before baiting}} \right\}$$

II- Biochemical Studies

The glassy clover snails *Monacha cartusiana* (Müller) were collected from clover fields in 2004/2005 season from Koom Hamada district and transferred to laboratories at Etay El-Broud Research Station, El-Behera governorate. The shell diameter of adult snails was measured at average 1.6 cm. They were acclimatized for a week under laboratory conditions and fed on lettuce (*Lactuca sativa*) *ad lib*. Toxic white bran baits were prepared containing each LC₅₀ or 0.5 LC₅₀ values of tested pesticides. The bait consisted of grain bran: molasses: blue dye with the ratio 8: 1: 1, respectively, (WHO, 1961). Baits were distributed in three replicates and 15 healthy animals were used in each replicate. In the case of control, pesticides were substituted with distilled water. Replicates were kept under laboratory conditions and suitable humidity required for snail activity. The dead animals were discarded, while the healthy adults were

taken at different time intervals, i.e. 1, 3, 5, and 7 days of treatment. The soft tissues were weighed at average (0.86 gm), pooled, and homogenized as 1: 10 (w/v) in buffer phosphate (0.1 M) PH 7.0. The homogenate was centrifuged at 5000 rpm for 20 min under cooling. The supernatant was decanted and frozen at -20 °C until used as enzymes source. Activities of some enzymes were measured such as alanine aminotransferases (ALT), aspartate aminotransferases (AST) according to **Retman and Frankel** method (1957). Additionally, acetylcholinesterase (AChE) activity was measured according to **Ellman et al** (1961) and its specific activity was expressed as µmole substrate hydrolyzed/mg protein/min. Total protein was measured according to **Lowry et al** (1951).

Statistical Analysis

In field treatments, analysis of variance was used to compare means among treatment. The least square means were compared for significant differences between treatments using student- Newman-Keuls test (**Sokal and Rohlf, 1969**). While, in biochemical studies all data were calculated as $\bar{X} \pm \text{S.E.}$ and comparison between two groups was performed by student t-test (**Motulesky, 1987**). P-value of 0.05 was considered significant.

RESULTS AND DISCUSSION

1. Molluscicidal activities of tested insecticides

The Survey and observation of the studied area showed that, glassy clover snails *Helix aspersa* (Müller) is the common species snails on winter crops. The

efficacy of tested pesticides under the field conditions was evaluated as snails' population reduction, bait consumption, days of pest control and the required poison baits (Kg/feddan). The results in **Table (1)** indicated that, methomyl and lufenuron were the most effective against brown snails causing (96.96 and 98.03%) and (85.44 and 94.66%) reduction in population for 0.5 LC₅₀ and LC₅₀, respectively. The population reduction by indoxacarb reached 79.02 and 93.33% with 0.5 LC₅₀ and LC₅₀, respectively. The lowest effective pesticide as molluscicide was Diple-2X, where the population reduction of snails reached 45.82 and 71.58% with the same concentrations, respectively.

Concerning the amounts of consumed baits, it was found that, Bt. bait exhibited the highest consumption by snails (7.6 Kg/feddan). While, the low consumptions were 2.5, 4.0, and 5.5 Kg/feddan for methomyl, lufenuron, and indoxacarb, respectively. On the other hand, the periods required to achieve successful control of the snails were 5, 12, 14, and 28 days for methomyl, lufenuron, indoxacarb and Diple-2X, respectively.

Generally, lufenuron and indoxacarb showed high molluscicidal effect against clover snails, which considered an important pest on most field crops, under field conditions compared with methomyl, in addition to other positive properties of the compounds low toxicity to non-target organisms and short persistence in the environment (Salagado, 1990; Harder, *et al* 1996 and Pluschkell *et al* 1998). Abou-El Khear *et al* (2005) found the same results under laboratory conditions. These results agree with the findings by Hussien (2003) who indicated that, methomyl was more effective as mollusci-

cidal compound using topical application and poisoned food techniques than indoxacarb. Abou-Baker (1997) illustrated that, the chemical control is the most powerful tool available for controlling snails. Zedan *et al* (1999) found that, bacterial formulation was the most effective against land snails compared with methomyl.

2. Toxicological activities of tested insecticides

Data on the Aspartate aminotransferases (AST), alanine aminotransferases (ALT), acetylcholinesterases activities and protein concentration in animals treated with 0.5 LC₅₀ and LC₅₀ of the tested pesticides at different time intervals are shown in **Tables (2 and 3)**. AST activity was reduced by 85.4 and 80.6% (Diple-2X); 69.6 and 53.3% (indoxacarb); 79.8 and 72.6% (lufenuron) and 63.6 and 67.7% (methomyl) for animals treated with 0.5 LC₅₀ and LC₅₀, respectively, after three days of treatment. Alanine aminotransferase was also significantly reduced compared with control animals. The inhibitory effects by 0.5 LC₅₀ and LC₅₀ for methomyl, lufenuron, indoxacarb, and Diple-2X on the AChE activity of snails are shown in **Table (3)**. The results indicated that, the highest inhibitory effect against AChE enzyme activity of treated animals was occurred with LC₅₀ of methomyl. The specific activity of AChE was expressed as $\mu\text{mole/mg protein/min}$ which accounted for 0.004, 0.004, 0.002, and 0.001 $\mu\text{mole/mg protein/min}$ for Diple-2X, indoxacarb, lufenuron, and methomyl, respectively. Slight inhibition in AChE enzyme activity was done with lufenuron and Diple-2X treatments. In addition, the protein

Table 2. Effect of tested pesticides against alanine aminotransferases (ALT), aspartate aminotransferases (AST) activities of glassy clover snails *Monacha cartusiana* (Müller) at different time intervals

Treatment	Enzyme Activity					
	Dose	Time interval (day)	AST (U / l)	% of Control	ALT (U / l)	% of Control
Control	-	1	47.3±7.5	100	18.7±2.4	100
		3	41.3±3.2	100	28.8±4.3	100
		5	43.0±3.1	100	26.8±4.6	100
		7	42.3±4.2	100	22.5±2.1	100
<i>Bt</i>	0.5LC ₅₀	1	25.7±4.6**	54.3	19.0±2.5	101.6
		3	35.0±4.8**	85.4	21.8±3.1**	75.6
		5	5.8±2.3**	13.5	12.0±3.2**	44.7
		7	7.7±1.3**	18.2	14.3±2.6**	63.5
	LC ₅₀	1	26.7±3.6 **	56.4	13.9±3.4**	74.2
		3	33.3±2.5 **	80.6	23.8±4.1**	82.3
		5	13.0±1.3**	30.2	18.7±3.6**	70.0
		7	7.73±2.1**	33.0	12.1±3.2**	53.8
indoxacarb	0.5LC ₅₀	1	22.0±3.5**	46.5	10.7±3.5**	57.2
		3	28.7±3.4**	96.6	12.1±1.3**	42.1
		5	5.7±1.2**	13.3	13.4±2.3**	50.0
		7	6.7±1.4**	28.7	22.2±1.8**	98.7
	LC ₅₀	1	33.0±5.2 **	69.7	9.1±2.1**	48.7
		3	22.0 ±1.2**	53.3	15.3±2.9**	56.8
		5	5.47±1.1**	12.7	12.1±3.7**	42.5
		7	7.47±1.4**	32.1	14.0±3.8**	62.2
lufenuron	0.5LC ₅₀	1	24.3±2.2*	51.3	13.2±2.4**	70.5
		3	34.3±3.2*	79.8	18.3±2.4**	63.6
		5	31.0±3.5	75.1	16.9±1.8**	63.3
		7	11.0±1.3**	47.2	19.6±3.7	87.5
	LC ₅₀	1	16.0±0.0**	33.8	12.9±1.7**	68.9
		3	30.0±0.8*	72.6	23.1±1.9*	80.2
		5	24.3±0.7**	56.5	20.2±2.3*	75.3
		7	15.0±0.2**	64.3	22.1±2.6	98.2
methomyl	0.5LC ₅₀	1	24.3±2.3**	51.4	19.3±3.1	103.2
		3	26.3±3.6**	63.6	14.2±1.6**	49.4
		5	4.5±1.1**	10.4	16.5±2.9**	61.2
		7	6.7±1.4**	28.8	14.9±2.9**	66.3
	LC ₅₀	1	27.3±7.5**	57.6	14.4±2.4**	48.3
		3	31.6±2.2**	76.7	17.0±3.5	93.7
		5	11.4±1.3**	26.5	14.0±1.8**	52.4
		7	3.2±0.0**	13.7	22.4±2.3	100

Each value is the mean of three replicates ± S. E.

* Significant at 0.05

** Significant at 0.01

Table 3. Effect of tested pesticides against acetyl cholinesterase (AChE) activity and total protein of glassy clover snails *Monacha cartusiana* (Müller) at different time intervals

Treatment	Dose	Time interval (day)	Enzyme Activity			
			AChE Activity ($\mu\text{mole/mg protein/min}$)	% of Control	Total protein (mg/ml)	% of Control
Control	-	1	0.002 \pm 0.0	100	120.6 \pm 2.5	100
		3	0.004 \pm 0.0	100	121.3 \pm 0.48	100
		5	0.003 \pm 0.001	100	128.6 \pm 3.8	100
		7	0.033 \pm 0.009	100	128.9 \pm 4.3	100
<i>Bt</i>	0.5LC ₅₀	1	0.003 \pm 0.00	67.23**	95.77 \pm 3.8**	78.51
		3	0.004 \pm 0.001	102.3	105.2 \pm 3.7*	86.65
		5	0.007 \pm 0.00	64.69**	96.67 \pm 2.8**	75.00
		7	0.023 \pm 0.001	36.20**	10.44 \pm 100*	77.88
	LC ₅₀	1	0.006 \pm 0.002	47.13**	28.00 \pm 5.6**	23.33
		3	0.005 \pm 0.001	95.83	77.00 \pm 4.6**	63.63
		5	0.004 \pm 0.00	57.24**	68.67 \pm 1.2**	53.42
		7	0.061 \pm 0.001	92.19	97.13 \pm 2.4	75.35
indoxacarb	0.5LC ₅₀	1	0.002 \pm 0.00	62.77**	65.0 \pm 4.9**	54.16
		3	0.005 \pm 0.001	62.69**	65.78 \pm 6.2**	54.36
		5	0.012 \pm 0.002	95.14	39.78 \pm 3.4**	30.93
		7	0.012 \pm 0.003	31.48**	99.65 \pm 4.6**	77.30
	LC ₅₀	1	0.009 \pm 0.003	68.98**	66.45 \pm 3.8**	55.09
		3	0.004 \pm 0.00	57.71**	59.78 \pm 5.7**	49.27
		5	0.023 \pm 0.012	58.06**	44.22 \pm 3.7**	34.46
		7	0.026 \pm 0.00	30.63**	76.09 \pm 1.3**	59.03
lufenuron	0.5LC ₅₀	1	0.003 \pm 0.00	69.82**	64.33 \pm 3.9**	53.34
		3	0.002 \pm 0.00	28.01**	56.33 \pm 4.6**	46.42
		5	0.003 \pm 0.00	39.64**	55.22 \pm 4.4**	42.9
		7	0.040 \pm 0.01	34.25**	82.00 \pm 5.3*	63.6
	LC ₅₀	1	0.002 \pm 0.00	58.87**	73.78 \pm 5.6**	61.17
		3	0.002 \pm 0.00	33.84**	87.78 \pm 2.5*	72.36
		5	0.003 \pm 0.00	63.85**	79.67 \pm 3.5**	61.95
		7	0.013 \pm 0.00	14.49**	73.00 \pm 7.5**	56.63
methomyl	0.5LC ₅₀	1	0.002 \pm 0.00	51.74**	81.78 \pm 3.2**	67.17
		3	0.004 \pm 0.00	63.30**	76.67 \pm 2.7**	61.25
		5	0.006 \pm 0.00	61.43**	53.33 \pm 2.5**	41.46
		7	0.028 \pm 0.01	23.53**	53.89 \pm 1.3**	41.46
	LC ₅₀	1	0.001 \pm 0.00	31.98**	74.33 \pm 4.6**	61.63
		3	0.003 \pm 0.00	60.75**	57.67 \pm 5.7	55.88
		5	0.004 \pm 0.00	47.00**	31.22 \pm 3.1**	47.60
		7	0.042 \pm 0.01	31.55**	46.56 \pm 1.1**	36.20

Each value is the mean of three replicates \pm S.E.

* Significant at 0.05

** Significant at 0.01

content was significantly decreased after methomyl, lufenuron, indoxacarb, and Diple-2X treatment. The highest reduction was recorded in the animals treated with the high concentration (LC₅₀) of methomyl. AChE was more vulnerable proteins than other enzyme system especially against methomyl that attributed to its toxic effect. Elevation of AST and ALT had been reported in pesticide treatments (El-Wakil *et al* 1992; Radwan *et al* 1992; Radwan *et al* 1993; and Mohamed, 1995). The possible mechanisms involved in the elevation of AST and ALT activities observed in the present study may be based on tissue damage, increased synthesis or decreased catabolism of both enzymes (Todior and Van-Heemastra, 1980). On the other hand, the decreased activity of AST and ALT may be due to rather leakage of the enzyme into extra cellular compartments or to actual enzyme inhibition by tested pesticides (Abou-Baker, 1997; Abdalla *et al* 1998 and Hussein, 2003).

REFERENCES

- Abdallah, E.M.A.; F.A. Kassem; H.B. El-Wakil and Y. Abou-Baker (1998). Molluscicidal potentially of several pesticides against *Eobania vermiculata* and *Theba pisena* terrestrial snails. *Annals Agric. Sci.* 17 (1): 263-276.
- Abou-Baker, Y.A. (1997). *Toxicological and Environmental Studies on Some Terrestrial Gastropods*. pp. 113-122. M. Sc. Thesis, Faculty of Agric. Univ. of Alexandria, Egypt.
- Abou-El Khear, R.K.; Kh.Y. Abdel-Halim and A.A. Hussien (2005). Molluscicidal efficiency of some pesticides against *Helix aspersa* land snails under laboratory conditions. *J. Agric. Sci. Mansoura Univ.*, 30(2): 1129-1134.
- Belfore, G.J.; R.K. Vadlamudi; Y.A. Osman and L.A. Bula (1994). A specific binding protein from tenebriomolitor for the insecticidal toxin of *Bacillus thuringensis* subsp tendnebrionis. *J. Bioch. & Biophys. Res. Comm.* 200 (1): 359-364.
- Ellman, G.L.; K.D. Country; V. Anders and R.M. Featherstone (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* 7: 88-95.
- El-Okda, M.M.K. (1980). Land snails of economic importance on vegetable crops at Alexandria and Neighboring regions. *Agric Res. Rev.* 58 (1): 79- 86.
- El-Okda, M.M.K. and K.A. Khalil (1981). Land mollusks economic pest attaching fruit orchards at Alexandria governorate. *Proc.4th Arab Pesticides Conf. Tanta Univ., Egypt.* 11: 289-302.
- El- Wakil, H.B.; M.A. Radwan and K.A. Osman (1992). Interaction of Some Oxime Carbamate Pesticides with Terrestrial *Helix aspersa* Snail Enzyme System. *Alex. Sci. Exch.*13 (3): 81-99.
- Flint, L. (1998). *Pests of the Garden and Snail Farm: A Grower's Guide to Using Less Pesticide*. 2nd Ed, Oakland: Univ . Calf. Div., Agric. and Nat. Resources Publication 3332.
- Godan, D. (1983). *Pest Slugs and Snails, Biology and Control*. pp. 191-192. Springer Verlag: Berlin.
- Harder, H.H.; S.L. Riley; S.F. Mocann and S.N. Irving (1996). Indoxacarb: A novel broad spectrum environmentally soft, insect control. *Proc. Brighton. Corp Protection Conf. Pests of Diseases, Brighton, UK*, pp. 449- 454.
- Henderson, C.F. and W. Tilton (1955). Tests with acaricides against the brown

- wheat mite. *J. Econ. Entomol.* **98**: 157–161.
- Hussien, A.A. (2003).** *Toxicological and Biochemical Studies on Some Economic Pests.* pp. 134-163. Ph. D. Thesis, Faculty of Agric., Univ. of Alexandria, Egypt.
- Kassab, A. and H. Daoud (1964).** Notes on the biology of land snails of economic importance in UAR. *Agric. Rev., Min. of Agric, Cairo*, **42**: 77-88.
- Lowry, A.M.; N.J. Rosebrough; A.L. Farr and R.J. Randall (1951).** Protein measurement with folin phenol reagent. *J. Biol. Chem.*, **193**: 256-275.
- Mohamed, Y.M. (1995).** *The Toxic Action of an Insecticide; Selecron 720 EC on Biological System.* pp.79-88. M.Sc., Thesis, Faculty of Agric., Cairo Univ., Egypt.
- Motulesky, H.I. (1987).** T-ease (tm), T-tests with ease, *ISI Software, Institute for Scientific Information.* Berkeley, CA.
- Nakhla, J.M.; A.W. Tadros; A.A. Abdel-Hafiz and A.G. Hashem (1993).** Survey and monitoring of land snails in pear orchards at the northern reclaimed lands. *Alex. Sci. Exch.* **14**(3): 43-57.
- Pluschkell, V.; A.R. Horowitz; G. Phllis and I. Ishaaya (1998).** DPX-MPO62 potent compound for controlling the Egyptian cotton leaf worm *Spodoptera littoralis* (Biosd). *Pestic. Sci.*, **54**: 85– 90.
- Radwan, M.A.; H.B. El-Wakil and K.A. Osman (1992).** Toxicity and biochemical impact of certain oxime carbamate pesticides against terrestrial snails *Theba pisana* (Muller). *J. Environ. Sci. Health, B* **27** (6): 759-773.
- Radwan, M.A.; K.A. Osman and A.K. Salama (1993).** Biochemical response of the brown garden snail *Helix aspersa* to chlorfluarzuron and flufenoxuron. *J. Environ. Health, B* **28** (3): 291 – 303.
- Retman, S. and S. Frankel (1957).** A colorimetric method for determination of serum glutamic oxaloacetic acid and glutamic pyruvic transaminase. *Am. J. Clin. Path.* **28**: 56– 63.
- Salagado, V.L. (1990).** Mode of action of insecticidal dihydro pyrazoles: selective block of impulse generation in sensory nerves. *Pestic. Sci.* , **23**: 389 –411.
- Sokel, R.R. and J.F. Rohlf (1969).** *Biometry.* Freeman, W.H–Co Stat / ANOVA Randomized Complete Blocks. San Francisco, USA.
- Todior, W.F. and E.A.H. Van-Heemastra (1980).** *Field Studies Monitoring Exposure and Effects in The Development of Pesticides.* pp. 207-213, Elsevier; Amsterdam.
- WHO, (1961).** Expert committee on bilharziasis, *Wid. Hilth., Org. Tech. Rep. Ser. No. 214.*
- Zedan, H.A.; A.A. Saleh and S.M. Abdail (1999).** Bacterial activity of *Bacillus thuringensis* against snails-toxicological and histological studies. *Proc.2nd Int. Conf. of Pest Control*, pp. 489-497 Mansoura, Egypt.

مجلة اتحاد الجامعات العربية للدراسات والبحوث الزراعية ، جامعة عين شمس ، القاهرة 14(2) ، 861-870 ، 2006 ،

الكفاءة الابدائية وسمية بعض المبيدات ضد قوقع البرسيم الزجاجي تحت الظروف الحقلية والمعملية

[56]

خالد يس عبد الحليم¹ - ربيع كامل أبو الخير² - علاء الدين عبد الفتاح حسين¹

1- المعمل المركزي للمبيدات - محطة البحوث الزراعية - وزارة الزراعة - ايتاي البرود - مصر

2- معهد بحوث وقاية النباتات-قسم الحيوانات الضارة والطيور-المركز الدولي للبطاطس-كفر الزيات- مصر

لأعلى إبادة هي (5 و 12 و 14 و 28 يوما
على التوالي).

* كانت كمية المبيد اللازمة لتحقيق المكافحة
الناجحة للقواقع 7.6 - 5.5 - 4 - 5.2
كجم /فدان للدابيل ، الافانت ، الماتش ثم
اللانيت على التوالي.

* أظهرت دراسة تأثير هذه المركبات على
نشاط بعض الإنزيمات الحيوية داخل جسم
القواقع المسممة أن التأثيرات المثبطة
للحلل من اللانيت والافانت على نشاط إنزيم
الاستيل كولين استريز كانت أعلى من
الماتش يليها الدابيل.

* حدث انخفاض واضح في نشاط إنزيمات
ALT, AST مما أدى الي تدهور النشاط
الجسمي للقواقع وقد حدث أعلى تثبيط
بعد ثلاثة أي-ام من المعاملة خاصة في
حالة استخ دام التركيز النصفى القاتل
LC₅₀.

أجريت هذه الدراسة تحت الظروف
الحقلية بمركز كوم حمادة محافظة البحيرة
ومحطة البحوث الزراعية بايتاي البرود
للموسم الزراعي 2004/2005 لدراسة كفاءة
ثلاث مركبات حديثة وأمنة بيئيا من مجاميع
مختلفة هي مركب الماتش (ليفينرون 15
%) (اندوكسكارب 5%) والمبيد البكتيري-
الدابيل (10.3 Bf). ومقارنتها بمركب اللانيت
(ميثومويل 90%) الموصى به في مكافحة
القواقع الأرضية.

تم استخدام تركيزين من هذه المركبات
تمثل التركيز النصفى قاتل LC₅₀ و نصف
هذا التركيز 0.5 LC₅₀ كطعوم سامة أظهرت
نتائج الدراسات الحقلية أن:-

* مركب اللانيت أعطى كفاءة ابدائية عالية
حتى 98% خلال خمسة أيام من المعاملة
يليه مركب الماتش (93,33%) ثم الافانت
(94,66%) وأخيرا مركب الدابيل لـ
(71,58%) وكانت المدة اللازمة للوصول

تحكيم: أ.د زيدان هندی عبد الحميد أ.د محمد عبد الهادي قنديل