

**SOME PHYSIOLOGICAL AND HISTOPATHOLOGICAL
EFFECTS OF TWO PESTICIDES AGAINST THE COTTON
LEAF WORM, *SPODOPTERA LITTORALIS* (BOISD.)**

[51]

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ABSTRACT

The toxicological effect of two pesticides, pyriproxyfen and abamectin were evaluated on third instar larvae of the cotton leafworm, *Spodoptera littoralis* (Boisd.). According to the estimated toxicity values, i.e. LC₅₀, LC₉₀ and slope values, the chemical abamectin showed that it was slightly more toxic than pyriproxyfen. The effect of these two insecticides, at the determined LC₅₀ and LC₂₅ values on the digestive physiology of treated larvae, as indicated by some nutritional indices was conducted. Obtained results demonstrated that pyriproxyfen and abamectin caused a significance reduction in growth of treated larvae as depicted by larval weight gain. Also, a significant decrease was found in the efficiency of ingested food to body matter and efficiency of conversion of digested food to body matter as compared to untreated insects. These effects were generally more evident when abamectin was used. The two tested compound also caused histological changes in the midgut of treated larvae, in form of disruption in the columnar epithelium cells and stretching leading to tearing in the peritrophic membrane. This observation might explain the impairment in nutritional indices in treated larvae as compared to the control.

Keywords: Cotton leafworm, Abamectin, Pyriproxyfen, Digestive indices, Midgut histology

INTRODUCTION

The cotton leafworm, *Spodoptera littoralis* (Boisd.) is considered the most serious pest infesting the Egyptian cotton plant. This insect also attacks other economically important crops such as; cucumber, potato, okra, egg plants and oth-

ers. In the past decade, the potency of different novel insecticides were used for the control of *S. littoralis*, e.g. benzoyl phenyl urea (chitin synthesis inhibitors), juvenile hormone mimics (hormones balance disrupters) and microorganism's fermentation derived compounds. Such agents have been looked at as selective

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agents to suppress both growth and development of larvae. Since then, several studies have been directed to elucidate the biochemical effects of such chemicals. For example, it was found that diflubenzuron, pyriproxyfen and abamectin caused a significant alteration of different enzymatic activities, i.e. esterase, chitinase and penoloxidase (Mostafa, 1993; Farag, 2001, Raymond-Delpech *et al* 2005). The literature cited on these compounds suggested their potential successful use as alternatives to classical insecticides. Furthermore such chemicals exhibit a relative short environmental persistence, low mammalian toxicity as well as the ease of their formulated application (Quistad *et al* 1974; Halley *et al* 1993; Dhadialla *et al* 1998).

The aim of the present study was to evaluate the toxicity of two insecticides pyriproxyfen and abamectin on a susceptible strain of the cotton leafworm *Spodoptera littoralis*. Also, the effect of these chemicals on nutritional indices of treated larvae was determined, as well, as their effect on the histological structure of the midgut.

MATERIAL AND METHODS

1- Biopesticides used

1-1- Pyriproxyfen: Admiral[®](10 % EC), is a juvenile hormone mimic (JHM), 4-phenoxyphenyl (RS)-2-(2-pyridyloxy) propyl ether. It was obtained from Sumitomo Chemical Co. (Osaka, Japan).

2-2- Abamectin: Vertimec[®] (1.8 % EC), belongs to the avermectins group (a microbial insecticides and acaricides) containing more than 80 % avermectin B1a and less than 20 % avermectin B1b. This

compound was obtained from Syngenta Agrochemical Co. (Qualiobia Governorate, Egypt).

2- Insect rearing technique

The experiments of the present work were carried out under laboratory conditions of 27 ± 2 °C and 65 ± 5 % R.H. The cotton leafworm, *S.littoralis* susceptible strain was obtained as egg-masses from Syngenta Agrochemical Co. (Qualiobia Governorate, Egypt). Each egg-mass was separately confined in sterilized jars, tapped with muslin covers. Upon larval hatching, fresh and clean castor-bean leaves were provided daily as food and clean jars were substituted for the used ones. At pupation, the pupae were sexed and then confined, in jars, at a sex ratio of 2 females to one male, until moth emergence. Upon adult emergence, moths were placed in glass globes and supplied with a cotton wick immersed in a 10 % sugar solution as a source of food. In addition two leaves of *Nerium oleander* were provided as an oviposition site. Deposited egg-masses were daily collected for further experiment.

3- Establishing toxicity lines

An emulsifiable formulation of each tested biopesticide was prepared in water as a stock solution and a series of six concentrations for each chemical were prepared from this solution, Clean castor bean leaves were sprayed with one of the prepared concentrations of each compound at rate of 0.7 ml/150 cm², the leaves were left to dry at room temperature.

Ten newly molted 3rd instar larvae, each with an average weight of 80 mg,

were placed in plastic cup measuring 20 cm diameter and provided with treated leaves with one of the prepared concentrations. The larvae were fed on treated castor bean leaves for two days and then offered untreated leaves. As a control, a similar number of larvae were provided with untreated castor bean leaves. Each concentration was replicated ten times. Mortality counts were assessed and corrected according to the formula of **Abbott (1925)**. Concentration mortality regression lines were carried out according to the method of **Finney (1972)** and the LC_{25} , LC_{50} and LC_{90} values determined.

4- Physiological Parameters

The digestive physiology as indicated by the calculation of nutritional indices were determined for third instar larvae treated with pyriproxyfen and abamectin at the determined LC_{25} and LC_{50} values, as well as their equivalent control.

For each treatment, 100 newly ecdysed (0-6 hours) 3rd instar larvae were placed in large plastic dishes. The initial fresh weight of larvae was determined before being offered a recorded weight of castor oil bean leaves treated with the calculated LC_{25} or LC_{50} of each of pyriproxyfen or abamectin for 48 hours. Larvae were then offered untreated leaves for the remaining duration of the larval stage. A control was set comprising an equivalent number of insects which were offered at all times untreated leaves.

The weights of larvae, the daily offered fresh castor oil leaves, faeces and unconsumed leaves of the previous day were recorded on a daily bases. The amount of diet eaten was estimated by the difference in dry weight of the diet before and after testing. From these records the

following nutritional indices were considered after 24, 72, 120, 168 and 240 hours following treatment, according to the method described by **Mcfarlane (1985)** and using the equations set by **Waldauer (1968)**.

- (1) Larval weight gain or loss.
- (2) Amount of ingested food.
- (3) Amount of digested food.
- (4) Approximate digestibility (AD).
- (5) Efficiency of conversion of ingested food (ECI) to body matter.
- (6) Efficiency of conversion of digested food (ECD) to body matter.

5- Histological study

To clarify the results of the nutritional indices in larvae treated with LC_{50} of either pyriproxyfen or abamectin, a histological study of the midgut was conducted. On the third day following treatment, 5 larvae from each treatment as well as from the untreated group were picked out and dissected. Their mid-guts were fixed in aqueous Bouin's solution, then dehydrated in ethanol solutions and cleared in xylene and then embedded in paraffin. Specimen were then cut at 6 μ thickness and then stained by Ehrlich's haematoxylin and counter stained by eosin.

6- Statistical analysis

Results were analyzed statistically using the Analysis of Variance (ANOVA) by the computer program (Sigma Plot version 2.0 for Windows) at a single time point which was examined by use of two-tailed student *t* test.

RESULTS

1- Toxicity values of pyriproxyfen and abamectin on *S. littoralis* larvae

According to toxicity regression lines (**Fig. 1**), both tested compounds exhibited an insecticidal activity to third instar larvae of *S. littoralis* (**Table 1**). Abamectin showed a higher toxic activity as expressed by the determined LC_{50} and LC_{90} , i.e. 69.11 and LC_{90} 239.11 ppm, respectively. Pyriproxyfen gave a higher values of LC_{50} 85.11 and LC_{90} 580.00 ppm. Both toxicity index and relative potency values based on the determined LC_{50} were 81.28, 1.23 for pyriproxyfen, 100.00 and 1.00 for abamectin, respectively. Also, the values of LC_{90}/LC_{50} ratio were 6.82 and 3.46 for pyriproxyfen and abamectin, respectively. The LC_{25} for these two respective mentioned chemicals was 42.5 and 23.91 ppm.

In addition, the slope value of abamectin was 2.38 higher than that of pyriproxyfen i.e. 1.58 (**Table 1**). Such difference in the slope values may be due to the slow action of pyriproxyfen compared to the relatively rapid action of abamectin (**Figure 1**).

2- Effect of pyriproxyfen and abamectin on nutritional indices

As shown in **Figure 2**, the larval growth factor as estimated by larval body weight gain post treatment was significantly affected in larvae treated with the two tested insecticides. Pyriproxyfen at its LC_{50} value of 85.11 ppm caused a lower weight gain in treated larvae by nearly 73, 72, 70, 48 and 30 % than their control, after 24, 72, 120, 168 and 240 hours, respectively. When LC_{25} (i.e. 42.50 ppm) of this chemical was tested,

the weight decrease in treated larvae than control larvae was 59, 62, 56, 57 and 46 % to the respective times mentioned. Abamectin caused a significantly higher reduction in larval weight than their equivalent control which was less by 45, 43, 36 and 25 % when LC_{50} (69.18 ppm) was tested and by 88, 85, 55, 55 and 52 % when LC_{25} (239.11 ppm) was used after the intervals of 24, 72, 120, 168 and 240 hours following treatment, respectively, (**Table 2**).

The ingested food parameter in larvae fed on castor oil leaves treated with either LC_{25} of pyriproxyfen or with both tested LC 's of abamectin was half that of their equivalent control, (i.e. 50%). Meanwhile, it was only slightly higher reaching 62.5% with LC_{50} of pyriproxyfen. The amount of digested food was also lowered by 60 and 40% than the control, as a result of treatment with LC_{50} values of pyriproxyfen and abamectin, respectively. However, it was 80% for both chemicals at LC_{25} values. The weight of unconsumed treated leaves by larvae was non-significantly higher reaching 108.33 and 101.66% after treatment with LC_{50} and LC_{25} of pyriproxyfen. Meanwhile, a significant 256.66% increase in unconsumed leaves was found after treatment with LC_{50} of abamectin (**Fig. 2**).

3- Histological effect:

The histological structure of the midgut in *S. littoralis* larvae show that it is lined by the peritrophic membrane which envelops the food particles followed by a closely arranged columnar epithelial layer with a rather broad apex, bearing a microvillus border. Also, observed are small goblet cells and both types of cells rest on

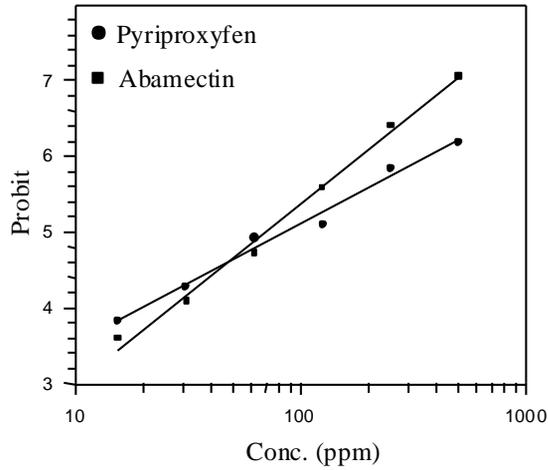


Fig. 1. Toxicity regression lines of pyriproxyfen and abamectin against third instar larvae of the cotton leafworm, *Spodoptera littoralis* (Boisd.)

Table 1. Toxicity values of pyriproxyfen and abamectin on the third instar larvae of *Spodoptera littoralis*

Compound	LC ₅₀ (ppm)	LC ₉₀ (ppm)	Slope	Toxicity index (Ti) based on LC ₅₀	Relative potency based on LC ₅₀	LC ₉₀ /LC ₅₀ ratio
Pyriproxyfen	85.11	580.00	1.58	81.28	1.23	6.82
Abamectin	69.18	239.11	2.38	100.0.	1.00	3.46

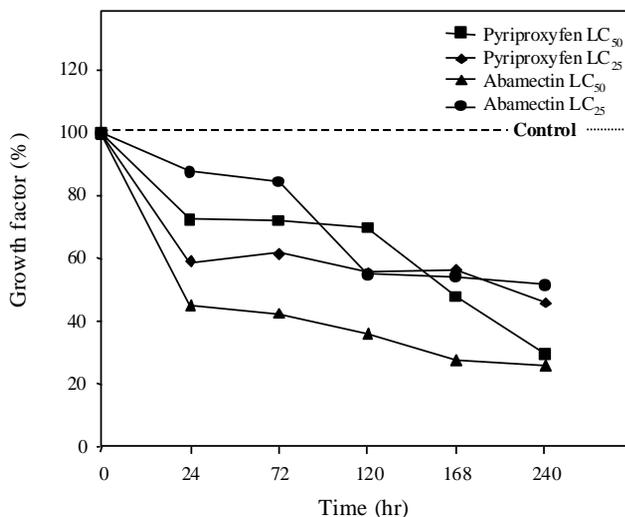


Fig. 2. Effects of pyriproxyfen and abamectin at LC₅₀ and LC₂₅ values on larval weights of third instar larvae of *Spodoptera littoralis* (Boisd.)

Table 2. Effect of pyriproxyfen and abamectin on nutritional indices of third instar larvae of the cotton leafworm, *Spodoptera littoralis* (Boisd.)

Nutritional index	chemical and tested concentration				
	Control	Pyriproxyfen		Abamectin	
		LC ₅₀	LC ₂₅	LC ₅₀	LC ₂₅
Weight of Ingested food (gr)	100.00 ± 0.003	62.50* ± 0.004	50.00* ± 0.003	50.00* ± 0.001	50.00* ± 0.002
Weight of unconsumed food (gr)	100.00 ± 0.010	108.33 ± 0.006	101.66 ± 0.006	256.66** ± 0.017	93.33 ± 0.003
Weight of digested food (gr)	100.00 ± 0.003	60.00* ± 0.004	80.00 ± 0.003	40.00** ± 0.001	80.00 ± 0.002
Weight of faeces t (gr)	100.00 ± 0.002	63.64* ± 0.001	63.64* ± 0.001	27.27*** ± 0.001	45.45** ± 0.001
ECI	100.00 ± 2.502	50.61* ± 3.36	61.02* ± 10.697	17.92*** ± 1.617	79.17 ± 21.89
ECD	100.00 ± 4.46	51.42* ± 1.743	53.55* ± 7.048	13.99*** ± 2.067	62.86* ± 1.065

Each value represents the mean of 100 larvae + Std. Dev. Compared to the control value, (***) highly significant $p \leq 0.001$, (**) moderately significant $p \leq 0.01$ and (*) significant $p \leq 0.05$ (student *t*-test).

ECI: Efficiency of conversion of ingested food to body matter.

ECD: Efficiency of conversion of digested food to body matter.

Fig (3a): Part of a cross section in the midgut of untreated 3rd instar larvae of *S. littoralis* [10 X].

- a) longitudinal muscles
- b) circular muscles
- c) basement membrane
- d) epithelium layer
- e) peritrophic membrane
- f) lumen

Fig (3b): Cross section in 3rd instar *S.littoralis* larvae treated with LC50 pyriproxyfen [3.2 X].

- e) basement membrane [broken up in some areas]
- d) epithelium layer [where the epithelial cells were detached from the basement membrane]
- e) peritrophic membrane
- g) cell debris

Fig(3c): Section in the midgut of 3rd instar *S. littoralis* larvae treated with LC50 abamectin [3.2 X].

- a) longitudinal muscles
- c) basement membrane [appears damaged]
- d) epithelium cells are disintegrated
- e) peritrophic membrane

a basement membrane. These layers are encircled by the muscular layers (**Fig. 3a**).

Some histological changes were observed in the midgut sections of larvae; prepared 3 days post treatment with LC₅₀ of pyriproxyfen in their diet. The distal ends of some columnar epithelial cells become somewhat distended with disrupted microvilli, while in other areas the cells remain intact, (**Fig. 3b**). The peritrophic membrane appears impaired and may be stretched or broken down in some places.

Application of abamectin at LC₅₀ value to the diet of third instar larvae caused a marked higher effect in the midgut of treated larvae (**Fig. 3c**). The midgut epithelium cells loose their columnar structure, in some areas these cells appear sloughed and the midgut lumen was filled with debris, presumably from degenerating cells. Only remnants of the peritrophic membrane can be detected.

DISCUSSION

The slow action of pyriproxyfen may be attributed to its long lasting mode of action as juvenile hormone mimics interfere with the hormonal balance at the molting process as mentioned by **Dhadialla et al (1998)**. Meanwhile, the relative rapid action of abamectin may be due to its mode of action as a neurotoxic insecticide which affect the γ -amino butyric acid (GABA) receptor and block the GABA-gated chloride channel in the affected larval brain (**Bloomquist, 1996** and **Raymond-Delpech et al 2005**). The reduction in ingested, digested and unconsumed food after treatment may be attributed to the effect of the tested chemicals on the activity of invertase, trehalase

and other digestive related enzymes as determined by **Rizk, (1998)**.

The obtained histological changes in the mid-gut of treated larvae are similar to results reported by **Mostafa and El-Attal, (1985)** in *S. littoralis* and (**Mohammed et al 1993**) in *Earias insulana*. Such effects were reported to be as a result of the tested compounds on stimulation of the cellular differentiation in mid-gut epithelia and formation of large autophagic vacuoles. In addition it was found that abamectin and pyriproxyfen caused several tissue and cell impairment which was characterized by hypertrophy of the epithelium. Similar results were reported in other insects treated with similar compounds, i.e. fire ant, *Solenopsis invicta*, cat flea, *Ctenocephalides felis*, *Pectinophora gossypiella*, and *Earias insulana* (**Glancey et al 1982; Meola et al 1996 & 2001**)

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بعض التأثيرات الفسيولوجية والنسجية المرضية لمبيدات حيويان على حشرة دودة ورق القطن (*Spodoptera littoralis* (Boisd.))

[51]

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وزن البراز اليرقي. أيضاً، فقد أمكن تقدير بعض القياسات الفسيولوجية والتي تمثلت في كفاءة تحويل الغذاء المتناول والمهضوم إلى مكونات جسمية. وقد أشارت النتائج المتحصل عليه إلى أن مبيد البيريبروكسيفين والأباميكيتين قد تسببا في إحداث خفضاً معنوياً في معامل النمو اليرقي ونقصاناً معنوياً في كفاءة تحويل الغذاء المتناول والمهضوم إلى مكونات جسمية. إضافة إلى ما سبق، فقد وجد أن كلا المبيدات قد تسببا في إحداث تغييرات نسيجية بمنطقة المعى الأوسط بيرقات العمر الثالث والتي قد تمثلت في إنحلال لطبقة الخلايا الطلائية وهو الأمر الذي أدى إلى حدوث انفصال للغشاء حول الغذائي بالمعى الأوسط.

تم تقوي م كل م من التائي مرات التوكسيكولوجية والفسيولوجية لمبيدات حيويان وهما البيريبروكسيفين والأباميكيتين وذلك على الطور اليرقي الثالث لحشرة دودة ورق القطن (*Spodoptera littoralis* (Boisd.)). وتبعاً لها تم الحصول عليه من نتائج سمية هذان المبيدات من قيم التركيزات المميتة لـ 50 و 90% وكذلك في م ميل لخطوط السمية، فقد تبين أن الأباميكيتين أكثر سمية من البيريبروكسيفين. أيضاً، فقد أمكن إختبار كل من التركيزات المميتة لـ 25 و 50% من هذه المبيدات على العديد من القياسات الفسيولوجية لهذه الحشرة. أمكن قياس كسل من وزن الغذاء المتناول، الغذاء المهضوم، الغذاء غير المتغذى عليه،

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