



## IN-VITRO EVALUATION OF PROBIOTIC BACTERIA SUPPLEMENTATION TO RUMINANT RATIIONS

[31]

El-Nagar<sup>1\*</sup>, S.E., Shemeis<sup>1</sup>A.R., Gouda<sup>1</sup>, G.F., El-Garhi<sup>2</sup>, M.S., Ebeid<sup>3</sup>, H.M., Azzaz<sup>3</sup>, H.H., Abdelgawad<sup>3</sup>, R.M.A., Mona, S. Zayed<sup>4</sup> and El-Bordeny<sup>1</sup>, N.E.

1. Animal Production Dept., Fac. of Agric., Ain Shams Univ., P.O. Box 68 Hadayek Shoubra, 11241, Cairo, Egypt.
2. Animal Reproduction Research Institute, Agricultural Research Center, Giza, Egypt.
3. Dairy Sciences Dept., National Research Centre, Dokki, Giza, 12311, Egypt
4. Agric. Microbiology Dept., Fac. of Agric., Ain Shams Univ., P.O. Box 68 Hadayek Shoubra, 11241, Cairo, Egypt

\*Corresponding author: [salama201013@agr.asu.edu.eg](mailto:salama201013@agr.asu.edu.eg)

Received 8 December, 2018,

Accepted 6 January, 2019

### ABSTRACT

The aim of this study was to evaluate effect of different level of probiotic supplementation to ruminant rations, using *in-vitro* batch culture technique to determine degradation and fermentation parameters. *In vitro* experimental ration was formulated, the ration consisted of 40% alfalfa hay and 60% concentrate feed mixture. Three level of probiotic supplementation ( $10^6$ ,  $10^8$ ,  $10^{10}$  cfu/kg DM) were evaluated. DM and total gas production as well as fermentation parameters of the incubated samples were determined after 24 hrs. of fermentation. Slightly increases ( $P>0.05$ ) in *in-vitro* dry matter degradability were observed for the ration supplemented with probiotics bacteria at different levels ( $10^6$ ,  $10^8$  and  $10^{10}$  cfu/ kg DM) compared to control ration. Probiotics bacteria supplementation with different level ( $10^6$ ,  $10^8$  and  $10^{10}$  cfu/ kg DM) led to significant ( $P<0.001$ ) increases in organic matter degradability and total gas production per sample and per g DM, OM, NDF and ADF compared to the not supplemented ration (control ration), and no significant differences were observed among the different levels of probiotics supplementation. Significant increase in total volatile fatty acid concentration after 24 hours' incubation period compared to the not supplemented ration. On the other hand, the treatment supplemented with probiotic recorded lower ammonia concentration compared to the control group. It could be concluded that, adding

probiotics bacteria supplementation to experimental ration resulted increase DM and OM degradability and using dose  $10^6$  CFU/kg DM feed is sufficient to induce improvement in degradability and fermentation parameters.

**Keywords:** *in-vitro*, probiotic, ruminant, fermentation.

### INTRODUCTION

Enhancement of animal productivity, efficiency of feed utilization and animal health are the main goal of rumen microbial studies. These aims could be achieved by producing a desirable fermentation product as probiotics or direct fed microbial (DFM). Many of the feed additives have been used to improve animal productivity and feed utilization efficiency. The probiotics are microbial growth promoters that could be manipulating the rumen fermentation characteristics in intestinal tracts of livestock animals (Weiss et al 2008).

The name probiotic comes from the Greek 'pro bios' which means 'for life'. The term "probiotic" has been defined as "a live microbial feed supplement, which affects beneficially of the host animal through improving the microbial balance in the intestine" (Fuller, 1989). Also, they are known as direct-fed microbial (DFM). Probiotic or DFM have been used to describe viable microorganisms, culture extracts, enzymes, exopolysaccharides or

various combinations of them (Yoon and Stern, 1995).

The use of probiotic additives has been developed as alternatives to antibiotics to improve animal health and productivity (Allen et al 2013). Probiotic supplements were also shown to increase carcass output and water holding capacity, and decrease cooking loss and meat hardness (Ceslovas et al 2005). *Lactobacillus bacillus* as a probiotic has several potential benefits like growth promotion of farm animals (Tripathi and Karim, 2009), protection against pathogens (Casas and Dobrogosz, 2000), alleviation of lactose intolerance (Mustapha and Savaiano, 1996), relief of constipation, antic-cholesterolemic effect, reduction of gut pH by stimulating the lactic acid producing microflora, competition with pathogens for a viable nutrient (Edens, 2003) and immunomodulation (Aottouri et al 2002).

The objectives of this study were to compare the effect of different levels of probiotic supplementation to ruminant ration on in-vitro degradation and fermentation parameters.

## MATERIAL AND METHODS

### Probiotic bacteria

### Microbial strains and growth condition

The probiotic bacteria used in this study is a mixture of 15 isolate of *Lactobacillus sp.* Lactobacilli isolates were grown on MRS broth (Oxoid) and Streptococci isolates were grown on M17 broth (Difco), after that the broth media incubated for 24 h at 37 °C. The strains were activated two or three times in order to obtain high biomasses in the stationary phase

### Experimental ration and treatments

*In-vitro* experimental ration was formulated; the tested ration contains 60:40 concentrate: roughage ratio. The CFM consisted of 60.89 % corn, 27.13 % soybean, 8.23 % flaxseed, 0.79% limestone 0.99 % sodium bicarbonate, 0.59 di-calcium phosphates, 0.40 trace premix and 0.79 salt. The data of chemical composition of the feed ingredients and tested rations are presented in Table (1). Four level of probiotic bacteria supplementation were applied 0, 10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU /kg DM of the tested ration.

**Table 1.** The chemical composition of the feed ingredients and tested rations

Item	alfalfa	Concentrate feed mixture
Dry matter	889.5	890.9
Organic matter	878.7	933.7
Neutral detergent fiber	460.6	184.3
Acid detergent fiber	359.7	59.4
Acid detergent lignin	41.6	10.4
Crude protein	208.5	157.3
Ether Extract	28.4	47.4
Ash	121.3	66.3
Non-fiber carbohydrate	181.2	544.7

### *In-vitro* gas production technique

Two days before beginning of the experiment, 400 (240 mg concentrate +160 mg alfalfa hay) ± 4 mg of sample for each treatment was weighed into 125 mL glass bottles. These bottles have a total volume of 125±2 mL. A buffer solution was prepared before addition of rumen fluid as described by McDougall (1948) and flushed continuously with CO<sub>2</sub> at 39°C during sample inoculation. Rumen fluid was obtained from slaughter house and it was collected from beef steers. The collected rumen fluid was mixed into a bottle (1L) with an O<sub>2</sub>-free headspace and immediately transported to laboratory at 39°C. Upon arrival at the laboratory, the rumen fluid was filtered through four layers of cheesecloth to eliminate large feed particles. The buffer solution was added to rumen fluid at ratio 4:1. forty mL of this inoculum was added to each bottle, then the headspace of each bottle was flushed with CO<sub>2</sub>, and closed. The initial pH of the inoculums was from 6.8-6.9. Triplicates of each sample were used for each treatment.

### Degradability

Dry matter degradability (% DMD) was calculated as the (difference between the sample DM content and that in the residual after 48 h incubation / sample DM content \* 100).

### Total gas production

After 24 h of samples incubation, the total gas production was estimated by the displacement of syringe piston, which was connected to the serum flasks. The gas produced due to fermentation of

substrate was calculated by subtracting gas produced in blank vessels (without substrate) from total gas produced in the vessels containing buffered rumen fluid and substrate.

**Calculation**

*In-vitro* organic matter digestibility (OMD, g/kg OM) were estimated according to (Menke and Steingass, 1988) as:

$$OMD = 14.88 + 0.889 GP + 4.5 CP (\%) + 0.0651 \text{ ash} (\%)$$

where GP is net GP in mL from 200 mg of dry sample after 24 h of incubation

After 24 hr of incubation, the filtrated rumen liquor for each sample was subjected for further investigation. The pH of rumen fluid was measured (pH meter) and quantitative analysis of ammonia concentration was carried out by Nesler method modified by Szumacher-Strabel et al (2002). Total volatile fatty acids (TVFA's) (Barnett and Reid, 1957).

**Gas production calculation**

After 24 hours' gas production was calculated as followed

- GPDM= total gas production (ml)/ substrate DM (g)
- GPOM= total gas production(ml)/ substrate OM (g)
- GPNDF= total gas production (ml)/ substrate NDF (g)
- GPADF= total gas production(ml)/ substrate ADF (g)

**Chemical analysis of feed ingredients**

Ration ingredients were analyzed for DM and ash, Crude fiber (CF); Crude protein (CP) (Nitrogen x 6.25) and ether extract (EE) contents according to AOAC (1997). Neutral detergent fiber (NDF), acid detergent fiber (ADF) and (ADL) acid detergent lignin contents were analyzed sequentially (Van Soest et al 1991) using the Ankom<sup>200</sup> Fibre Analyzer for NDF and ADF. The NDF content was analyzed with 2 additions of heat-stable α-amylase and 1:1 g sodium sulfite per g sample in the neutral detergent solution. NDF and ADF are expressed inclusive of residual ash. Non-fiber carbohydrate (NFC) was calculated according to the following formula:  
 $NFC(\%) = 100 - (\%ND + \%CP + \%fat + \%ash)$  (NRC, 2001).

**Statistical analysis**

The data of *In-vitro* degradability and fermentation parameters were statistically analyzed according to statistical analysis system User's Guide, (SAS, 1998). Separation among means was carried out by using Duncan Multiple test, (Duncan, 1955). The following model was used:

$$Y_{ij} = \mu + T_i + e_{ij}$$

Where: Y<sub>ij</sub> = the observation of the model, μ = General mean common element to all observation, T<sub>i</sub> = the effect of the treatment i, and e<sub>ij</sub> = the effect of error

**RESULTS AND DISCUSSION**

**Dry matter and organic matter degradability**

The data of Table (2) showed Effect of probiotics supplementation doses (0, 10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) on *in-vitro* dry matter and organic matter degradability. The data clearly showed that, slightly (P>0.05) increases in *in-vitro* dry matter degradability were observed for the experimental ration supplemented with probiotics bacteria at different levels (10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) compared to control ration (not supplemented). The heights dry matter degradability was recorded for level of 10<sup>6</sup> CFU/ kg DM (46.45 g/kg) followed by level 10<sup>10</sup> CFU/ kg DM (45.77 g/kg) then 10<sup>8</sup> CFU/ kg DM (43.48 g/kg), while the lowest value was recorded for control (not supplemented) (43.21 g/kg).

**Table 2.** Effect of probiotics supplementation doses (0, 10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) on *in-vitro* dry matter and organic matter degradability (DMD and OMD).

Degradation	control	Probiotic level CFU/kg DM			SE	P value
		10 <sup>6</sup>	10 <sup>8</sup>	10 <sup>10</sup>		
Dry matter, %	43.21	46.45	43.48	45.77	1.13	0.245
Organic matter, %	33.97 <sup>b</sup>	36.53 <sup>a</sup>	36.04 <sup>a</sup>	36.00 <sup>a</sup>	0.37	0.001

Different superscript are significantly different (P<0.05)

On the other hand, Probiotics bacteria supplementation with different level (10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) led to significant (P<0.001) increases in organic matter degradability (%) compared to the not supplemented ration (control ration) (Table,

2), moreover no significant differences were observed among the different levels of probiotics supplementation ( $10^6$ ,  $10^8$  and  $10^{10}$  CFU/ kg DM). These may be due to the probiotic supplementation which stimulate rumen bacteria growth (**Chiquette et al 2008**) and fermentation (**Stein et al 2006**), consequently improve DM degradation. The highest OM degradability was recorded for level of  $10^6$  CFU/ kg DM (36.53 g/kg) followed by  $10^8$  CFU/ kg DM (36.04 g/kg) then  $10^{10}$  CFU/ kg DM (36.00 g/kg), while the lowest value was recorded for control (33.97 g/kg). The data point to that it could be used the probiotics at level of  $10^6$  CFU/ kg DM.

These results are in line with the earlier report of **Sheikh et al (2017)** when add probiotic mix contains *Saccharomyces* and *Lactobacillus acidophilus* to the ration which found increase in DM and OM degradability as well as gas production compared to control. Also **Ganai et al (2015)** recorded higher *in-vitro* DM and OM digestibility values at supplementation of yeast to bajra straw based complete ration using goat rumen liquor. **Malik and Singh (2009)** also reported improvement in *in-vitro* or *in sacco* degradability pattern of nutrients due to supplementation of yeast culture.

### Gas production

Gas production is a good indicator of microbial ferment ability, digestibility and rumen protein production (**Salem et al 2014**). *In-vitro* gas production per g dry matter (GP/g DM), organic matter (GP/g OM), degraded dry matter (GP/g dDM), degraded organic matter (GP/g dOM), neutral detergent fiber (GP/g NDF) and acid detergent fiber (GP/g ADF) after 24 hours' incubation period as a response to increasing probiotics bacteria supplementation level (0,  $10^6$ ,  $10^8$  and  $10^{10}$  CFU/ kg DM) to the experimental ration are presented in **Table (3)**. Probiotics bacteria supplementation with different level ( $10^6$ ,  $10^8$  and  $10^{10}$  CFU/ kg DM) resulted significant increases in *in-vitro* total gas production per sample and per g DM, OM, NDF and ADF after 24 hours' incubation period compared to the not supplemented experimental ration (control ration). While, no significant differences were observed among the different levels of probiotics supplementation ( $10^6$ ,  $10^8$  and  $10^{10}$  CFU/ kg DM). This increase in total gas accumulation may be attributed to effect of probiotic that led to increase in OM degradability (table 2). These results are agree with **Sheikh et al (2017)** who found increase in total

gas production when add probiotic mix contains *Saccharomyces* and *Lactobacillus acidophilus* to the ration compared to control. Also **Ganai et al (2015)** recorded higher *in-vitro* total gas production when supplemented bajra straw based diet with yeast. In this connection **Blümmel and Ørskov (1993)** reported that fermentation of organic compounds produces gas as one of the end-products providing the foundation of the strong correlation between OM digestibility and volume of gas produced.

Also significant increase was observed in *in-vitro* total gas production per g dDM was recorded for level of  $10^8$  CFU/ kg DM compared to the control ration, while both treatment not significantly differed with  $10^6$  and  $10^{10}$  CFU/ kg DM. On the other hand, no significant differences were observed among the different experimental ration in total gas production (ml) per g dOM (**Table 3**).

### Fermentation parameters

*In-vitro* fermentation parameters pH value, ammonia and volatile fatty acids (VFA's) concentration after 24 hours' incubation period with increasing probiotics bacteria supplementation (0,  $10^6$ ,  $10^8$  and  $10^{10}$  CFU/ kg DM) are presented in **Table (4)**. Probiotics bacteria supplementation with different level ( $10^6$ ,  $10^8$  and  $10^{10}$  CFU/ kg DM) resulted significant increase in total volatile fatty acid concentration after 24 hours' incubation period compared to the not supplemented ration. The highest VFA's

Concentration was recorded for level of  $10^6$  CFU/ kg DM (7.71 mg %) followed by  $10^8$  CFU/ kg DM (7.69 mg %) then  $10^{10}$  CFU/ kg DM (6.96 mg %), while the lowest value was recorded for control (6.04 mg %). These results may be due to effect of probiotic supplementation which led to improve degradability and total gas production as indicated in **Tables (2 and 3)** consequently led to increase rumen fermentation. Volatile fatty acids are the ultimate product of microbial fermentation in the rumen and they are the main source of metabolizable energy for ruminants (**Van Soest, 1982**).

On the other hand, the treatment supplemented with probiotic recorded lower ammonia concentration compared to the control group. This may be due to *lactobasillus sp* is the main strain in our probiotics which improve carbohydrate fermentation.

**Table 3.** Effect of probiotics supplementation doses (0, 10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) on *in-vitro* gas production as ml per g DM, OM, dDM, dOM, NDF and ADF after 24 hours' incubation period.

Total gas production	control	Probiotic level, CFU/kg DM			SE	P value
		10 <sup>6</sup>	10 <sup>8</sup>	10 <sup>10</sup>		
per sample	37.89 <sup>b</sup>	41.78 <sup>a</sup>	41 <sup>a</sup>	40.38 <sup>a</sup>	0.62	0.0008
GP/g DM, ml	104.68	115.30	112.78	111.54	1.7	0.012
GP/g dDM, ml	87.94 <sup>b</sup>	90.17 <sup>ab</sup>	94.32 <sup>a</sup>	87.30 <sup>b</sup>	1.75	0.037
GP/g OM, ml	114.98 <sup>b</sup>	126.65 <sup>a</sup>	123.88 <sup>a</sup>	110.47 <sup>a</sup>	5.71	0.195
GP/g dOM, ml	111.50	114.36	113.74 <sup>a</sup>	112.11 <sup>a</sup>	1.48	0.499
GP/ g NDF, ml	322.57 <sup>b</sup>	355.17 <sup>a</sup>	347.23 <sup>a</sup>	306.97 <sup>a</sup>	17.18	0.212
GP/ g ADF, ml	530.15 <sup>b</sup>	583.54 <sup>a</sup>	570.23 <sup>a</sup>	502.08 <sup>a</sup>	29.12	0.221

Different superscript are significantly different (P<0.05)

Probiotics bacteria supplementation with different level (10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) resulted significant reduction in pH value after 24 hours incubation period compared to the not supplemented experimental ration (control ration). These may be due to the effect of the probiotic supplementation on TVFA's and ammonia concentration (**Table 4**), which the pH is affected by TVFA's ammonia concentration.

**Table 4.** Effect of probiotics supplementation doses (0, 10<sup>6</sup>, 10<sup>8</sup> and 10<sup>10</sup> CFU/ kg DM) on in-vitro fermentation parameters after 24 hours' incubation period.

Item	control	Probiotic level, CFU/kg DM			SE	P value
		10 <sup>6</sup>	10 <sup>8</sup>	10 <sup>10</sup>		
pH	5.77 <sup>a</sup>	5.56 <sup>c</sup>	5.50 <sup>c</sup>	5.64 <sup>b</sup>	0.02	0.0001
Ammonia, mg/dl	14.42	13.196	12.85	12.96	0.61	0.2839
Volatile fatty acid, meq/dl	6.04 <sup>b</sup>	7.71 <sup>a</sup>	7.69 <sup>a</sup>	6.96 <sup>a</sup>	0.27	0.0028

Different superscript are significantly different (P<0.05)

**CONCLUSION**

It could be concluded that, adding probiotics bacteria supplementation to experimental ration resulted increase DM and OM degradability and using dose of 10<sup>6</sup> CFU/kg DM feed is sufficient to induce improvement in degradability and fermentation parameters.

**ACKNOWLEDGEMENTS**

All authors would like to express their thankful to The Science & Technology Development Fund (STDF), Ministry of Higher Education & Scientific Research, Egypt for providing financial support for this study through national challenges target project " Production of probiotics and evaluation its impact on dairy and beef cattle performance, ID-10802".

**REFERENCES**

Allen, S.J., Wareham, K., Wang, D., Bradley, C., Hutchings, H., Harris, W. and Mack, D., 2013. Lactobacilli and bifidobacteria in the prevention of antibiotic-associated diarrhoea and Clostridium difficile diarrhoea in older inpatients (PLACIDE): a randomised, double-blind, placebo-controlled, multicentre trial. *The Lancet*, 382(9900), 1249-1257.

AOAC, 1997. Association of Official Analytical Chemists. *Official Methods of Analysis*, vol. 16, 3<sup>rd</sup> Revision, Gaithersburg, MD, USA.

Aottouri, N., Bouras, M., Tome, D., Marcos, A., and Lemonnier, D., 2002. Oral ingestion of lactic-acid bacteria by rats increases lymphocyte proliferation and interferon-γ production. *British J. of Nutr.*, 87(4), 367-373.

Barnett, A.J.G., and Reid, R.L., 1957. Studies on the production of volatile fatty acids from the grass by rumen liquor in an artificial rumen. *J. Agric. Sci.*, 48, 315-321.

- Blümmel, M. and Orskov E.R., 1993. Comparison of gas production and nylon bag degradability of roughages in predicting feed intake in cattle. *Anim. Feed Sci. Technol.* **40**, 109-119.
- Casas, I.A. and Dobrogosz, W.J., 2000. Validation of the probiotic concept: *Lactobacillus reuteri* confers broad-spectrum protection against disease in humans and animals. *Microbial ecology in Health and Disease*, **12**(4), 247-285.
- Ceslovas, J., Vigilius J. and Almantas S., 2005. The effect of probiotic and phytobiotics on meat properties and quality in pigs. *Vet. Zootech.*, **29**, 80-84.
- Chiquette J., Allison, M.J. and Rasmussen, M.A., 2008. *Prevotella bryantii* 25a used as a probiotic in early-lactation dairy cows: Effect on ruminal fermentation characteristics, milk production, and milk composition. *J. of Dairy Sci.*, **91**, 3536-3543.
- Duncan, D.B., 1955. Multiple range and multiple F tests. *Biometrics*, **11**(1), 1-42.
- Edens, F.W., 2003. An alternative for antibiotic use in poultry: probiotics. *Revista Brasileira de Ciência Avícola*, **5**(2), 75-97.
- Fuller, R., 1989. Probiotics in man and animals. *J. of Appl. Bacteriology* **66**, 365-378.
- Ganai, A.M., Sharma, T. and Dhuria, R.K., 2015. Effect of yeast (*Saccharomyces cerevisiae*) supplementation on ruminal digestion of bajra (*Pennisetum glaucum*) straw and bajra straw-based complete feed in-vitro. *Anim. Nutr. Feed Technol.* **15**, 145-153.
- Jiang, T., Mustapha, A., and Savaiano, D.A. 1996. Improvement of lactose digestion in humans by ingestion of unfermented milk containing *Bifidobacterium longum*. *J. of Dairy Sci.*, **79**(5), 750-757.
- Malik R. and Singh, R., 2009. Effect of yeast and fungi culture on in-vitro ruminal fermentation. *Indian J. Anim. Nutr.* **26**, 40-45.
- McDougall, E. I., 1948. Studies on ruminant saliva. 1. The composition and output of sheep's saliva. *Biochemical Journal*, **43**(1), 99-109.
- Menke, K.H., Steingass, H., (1988). Estimation of the energetic feed value from chemical analysis and in-vitro gas production using rumen fluid. *Animal research and development*, **28**, 7-55.
- NRC, 2001. *Nutrient Requirements of Dairy Cattle*. 7<sup>th</sup> Rev. Ed. National Academy Press, Washington, DC, USA.
- Salem A.Z.M., Kholif, A.E., Elghandour, M.M.Y., Hernandez, S.R., Domínguez-Vara, I.A. and Mellado M., 2014. Effect of increasing levels of seven tree species extracts added to a high concentrate diet on in vitro rumen gas output. *Anim. Sci. J.*, **85**, 853-860.
- SAS., 1998. *Statistical Analysis System*. User's Guide Inst., Inc. Cary, NC, USA.
- Sheikh G.G., Ganai, A.M., Ishfaq, A., Afzal, Y. and Ahmad, H.A., 2017. In-vitro effect of probiotic mix and fibrolytic enzyme mixture on digestibility of paddy straw. *Adv. Anim. Vet. Sci.* **5**(6), 260-266.
- Stein, D.R., Allen, D.T., Perry, E.B., Bruner, J.C., Gates, K.W., Rehberger, T. G., and Spicer, L.J., 2006. Effects of Feeding Propionibacteria to Dairy Cows on Milk Yield, Milk Components, and Reproduction1. *J. of Dairy Sci.*, **89**(1), 111-125.
- Szumacher-Strabel, M., Potkanski, A., Kowalczyk, J., Cieslak, A., Czauderna, M., Gubala, A. and Jedroszkowiak, P., 2002. Influence of supplemental fat on rumen volatile fatty acid profile, ammonia and pH level in sheep feed standard diet. *J. of Anim. Physiology and Nutrition Sci.*, **11**(4), 577-587.
- Tripathi M.K. and Karim S.A., 2009. Effect of individual and mixed live yeast culture feeding on growth performance, nutrient utilization and microbial crude protein synthesis in lambs. *J. of Anim. Feed Sci. and Technol.* **155**(2), 163-171.
- Van Soest, P.J., 1982. *Nutritional ecology of the ruminant*. Cornell University Press, Ithaca, NY, USA.
- Van Soest, P.V., Robertson, J.B., and Lewis, B.A., 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *Journal of dairy science*, **74**(10), 3583-3597.
- Weiss, W.P., Wyatt, D.J., and McKelvey, T.R., 2008. Effect of feeding propionibacteria on milk production by early lactation dairy cows. *Journal of Dairy Science*, **91**(2), 646-652.
- Yoon, I.K., and Stern, M.D., 1995. Influence of direct-fed microbials on ruminal microbial fermentation and performance of ruminants: A review. *Asian-Aust. J. Anim. Sci.*, **8**(6), 533-555.



## تقييم بكتيريا البروبيوتيك معمليا على علائق المجترات

[31]

سلامة السيد النجار<sup>1</sup> - احمد راغب شمس<sup>1</sup> - جودة فتحى جودة<sup>1</sup> - محمد سامى الجارحي<sup>2</sup> -  
حسام محروس عبيد<sup>3</sup> - حسام الدين حسين عزاز<sup>3</sup> - رمضان محمد أحمد عبد الجواد<sup>3</sup> - منى سعيد زايد<sup>4</sup> -  
نصر السيد البردينى<sup>1</sup>

1. قسم الانتاج الحيوانى - كلية الزراعة - جامعة عين شمس - ص.ب. 68 حدائق شبرا 11241 - القاهرة - مصر
2. معهد بحوث تناسليات - مركز البحوث الزراعية - الجيزة - مصر
3. قسم علوم الالبان - المركز القومى للبحوث - الدقى - الجيزة - 12311 - مصر
4. قسم الميكروبيولوجيا الزراعية - كلية الزراعة - جامعة عين شمس - ص.ب. 68 حدائق شبرا 11241 - القاهرة - مصر

\*Corresponding author: [salama201013@agr.asu.edu.eg](mailto:salama201013@agr.asu.edu.eg)

Received 8 December, 2018, Accepted 9 January, 2019

ومجموع إنتاج الغاز لكل عينة ولكل جرام مادة جافة ومادة عضويه والألياف الذائبة فى الوسط المتعادل والألياف الذائبة فى الوسط الحامض مقارنة بالعليقه الضابطة، ولم يلاحظ أى فروق معنويه بين المستويات المختلفة من البروبيوتيك. ويوجد زيادة معنويه فى إجمالي تركيز الأحماض الدهنيه الطياره بعد فترة تحضين 24 ساعه مقارنة بالمجموعة الضابطة، من ناحيه أخرى وجد إنخفاض تركيز الأمونيا فى المجموعة المعاملة بالبروبيوتيك مقارنة بالمجموعة الضابطة؛ يستنتج من ذلك أن إضافة بكتيريا البروبيوتيك للعلائق التجريبيه أدى إلى زيادة تحلل المادة الجافه والمادة العضويه وإستخدام الجرعة  $10^6$  وحدة خليه لكل كجم مادة جافه كافيه لتحسين التحلل ومقاييس التخمر.

الكلمات الداله: الهضم المعمل، البروبيوتيك، المجترات، التخمر

## الموجز

تهدف الدراسة لتقييم مستويات مختلفة من البروبيوتيك فى علائق المجترات، وذلك بإستخدام التخمر المعمل لتحديد مقدار التحلل ومقاييس التخمر، وتتكون العليقة من 40% دريس البرسيم الحجازى و60% مخلوط علف مركز، وكانت مستويات إضافة البروبيوتيك هى  $10^6$ ،  $10^8$ ،  $10^{10}$  خليه لكل كجم مادة جافه من العليقه، تم تقدير المادة الجافه وإنتاج الغاز الكلى وكذلك مقاييس التخمر بعد مرور 24 ساعه من التخمر، لوحظ زيادة غير معنويه ( $P > 0.05$ ) لتحلل المادة الجافة بإستخدام بكتيريا البروبيوتيك عند مستويات مختلفة ( $10^6$ ،  $10^8$ ،  $10^{10}$  وحدة خليه لكل كجم مادة جافه) مقارنة بالعليقه الضابطة، كما أدت إضافة بكتيريا البروبيوتيك بمستويات مختلفة ( $10^6$ ،  $10^8$ ،  $10^{10}$  وحدة خليه لكل كجم مادة جافه) إلى زيادة معنويه ( $P < 0.001$ ) فى تحلل المادة العضويه

تحكيم: ا.د عادل عيد محمود

ا.د محمود خورشيد